SYNTHESIS OF 3'-AMINO-3'-DEOXYADENOSINE FROM LEVOGLUCOSENONE

Hideyuki Kuno,* Shigeo Niihata, Takashi Ebata, and Hajime Matsushita

Life Science Research Laboratory, Japan Tobacco Inc., 6-2 Umegaoka, Aobaku, Yokohama, Kanagawa 227, Japan

<u>Abstract</u>-----A synthesis of 3'-amino-3'-deoxyadenosine (2) was developed by utilizing levoglucosenone (1) as a starting material through regio- and stereoselective *cis*-oxyamination of the carbon-carbon double bond.

Levoglucosenone [1,6-anhydro-3,4-dideoxy- β -D-glycero-hex-3-enopyranos-2-ulose (1)]¹ is widely known as a pyrolytic product of cellulose. It is a useful chiral source for synthesizing natural products² because of its highly functionalized structure which contains useful one chiral center. For several years, we have focused our attention mainly on its utilization for the synthesis of a variety of useful compounds as a chiral auxiliary.³ Attention is now being directed to new applications for synthesizing this nucleoside derivative.

3'-Amino-3'-deoxyadenosine (2) was isolated from the microbial culture filtrates, 4^{-7} which inhibited the activity of protein biosynthesis⁸ and showed remarkable antibiotic activity.^{7,8} On the other hand, it was also reported that the 5'-triphosphate of 3'-amino-3'-deoxyadenosine (2) effectively inhibited the DNA-dependent RNA polymerase by being incorporated into the 3' end of the growing RNA chain.⁹⁻¹¹ Because of the interesting bioactivity of 3'-amino-3'-deoxyadenosine (2), there have been several reports about the synthesis of 2 to date.¹²⁻¹⁹ Described here is a new approach to synthesize 3'-amino-3'-deoxyadenosine (2) with emphasis on the versatility of levoglucosenone (1) (Figure 1).

A practical synthesis is shown in Scheme 1. Compound (3) was easily prepared from levoglucosenone (1) in 2 steps by the reported procedure.^{2,3} The oxyamination of the carbon-carbon double bond of 3 with OsO4 and t-butyl *N*-chloro-*N*-sodiocarbamate under the Sharpless condition²⁰ afforded compound (4) as a single







Figure 2 An ORTEP drawing of 4.



Scheme 1

HETEROCYCLES, Vol. 41, No. 3, 1995

isolable isomer in 59.8 % yield. The structure of 4 was confirmed by X-ray diffraction analysis. Good crystals of 4 were obtained after purification and they were submitted for X-ray analysis. The ORTEP view of 4 is shown in Figure 2. It clearly indicates that the amino and hydroxy groups are introduced with high regio- and stereoselectivities in the oxyamination of 3. The $3.4-O_N$ -isopropylidene derivative (5a) was obtained by the reaction of 4 and 2,2-dimethoxypropane with p-toluenesulfonic acid. Treatment of 5a with tetrabutylammonium fluoride gave the alcohol (5b) in 62.7 % yield. The oxidation of 5b with oxalyl chloride and dimethyl sulfoxide under the Swern condition²¹ afforded 6 in 88.3 % yield. Based on the studies of our group, it had been found that the Baeyer-Villiger oxidation was effective for the transformation of the levoglucosenone skeleton to the γ -lactone.³ In this case, 7a and 7b were effectively obtained by the Baeyer-Villiger oxidation of 6 with monoperoxyphthalic acid magnesium salt. Furthermore, both 7a and 7b were converted to 8 by the reduction with diisobutylaluminium hydride. The acylation of 8 with acetic acid, acetic anhydride, and sulfuric acid as a catalyst, afforded 9 directly in 81.1 % yield. Tetraacetylaminoribofranose (9) was a mixture of the α - and β - forms, however, it was not necessary to separate their anomers for subsequent conversion into nucleosides by reaction with the adenine derivative. The coupling reaction of 9 and N⁶-benzoyl-bis(trimethylsilyl)adenine was performed in the presence of SnCl4/1,2-dichloroethane at 80°C¹⁹ to afford 10 as a single isomer in 55.1 % yield. After deprotection with methanolic sodium methoxide it afforded 3'-amino-3'-deoxyadenosine (2) in 85.2 % yield. The physical data of our synthetic 2 were identical with those reported. 15,16

In conclusion, a synthesis of 3'-amino-3'-deoxyadenosine (2) was achieved starting from levoglucosenone (1) in 11 steps and the overall yield was 3.3 %.

EXPERIMENTAL

The ir spectra were measured on a Jasco FT/IR-5000 photospectrometer. ¹H-Nmr spectra were recorded at 300 MHz on a Brucker AC-300P spectrometer. Optical rotations were measured on a Jasco DIP-370 polarimeter. Elemental analysis was performed on a Yanagimoto CHN recorder MT-3. Melting points were measured with a Yanagimoto melting point apparatus.

1,6-Anhydro-2-t-butyldiphenylsilyl-4-deoxy-4-t-butylcarbamide- β -D-altropyranose (4).

To a solution of *t*-butyl carbamate (3.51 g, 30.0 mmol) in methanol (20.0 ml), *t*-butyl hypochlorite (3.39 ml, 30.0 mmol) was carefully added dropwise at 0°C, after 15 min, a methanolic solution (40.0 ml) of sodium

hydroxide (1.26 g, 31.5 mmol) was added dropwise, and the reaction mixture was stirred at room temperature for 1 h. The solvent was removed to give the crude t-butyl N-chloro-N-sodiocarbamate as a white solid. Addition of acetonitrile (80.0 ml) and AgNO3 (5.21 g, 30.0 mmol) resulted in the gradual appearance of a brown suspension. The reaction mixture was stirred at room temperature for 30 min, and then 1,6-anhydro-2*t*-butyldiphenylsilyl-3,4-dideoxy- β -D-threo-hexo-3-enopyranose (3)³f) (7.21 g, 20.0 mmol), 1M OsO4/*t*-butyl alcohol (2.0 ml, 2.0 mmol), and water (1.7 ml) were added. The reaction suspension was stirred at room temperature for 3 d. Filtration of the reaction mixture through Celite gave a yellow-brown solution. The filtrate was refluxed for 3 h with 5% aqueous sodium sulfite (50.0 ml). The resulting mixture was concentrated and extracted with dichloromethane three times. The organic layer was dried over anhydrous magnesium sulfate and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (n-hexane:ethyl acetate=3:1). Furthermore, crystallization from the mixture of *n*-hexane and ethyl acetate afforded 4 (5.87 g, 59.8 % yield); mp 170-171.5°C; $[\alpha]^{25}$ -54.3° (c 0.56, CHCl₃); ¹H-nmr (CDCl₃) δ: 7.75-7.70 (4H, m, Ar-H), 7.48-7.36 (6H, m, Ar-H), 5.01 (1H, d, J=1.6 Hz, H-1), 4.78 (1H, br d, N-H), 4.52 (1H, m, H-5), 4.05 (1H, m, H-3), 3.98(1H, m, H-4), 3.84-3.73 (2H, m, H-6), 3.49 (1H, dd, J=8.2, 1.6 Hz, H-2), 2.07 (1H, br d, O-H), 1.42 (9H, s, t-Bu), 1.09 (9H, s, t-Bu); Ir (KBr) 1700 (s), 1508 (s), 1429 (m), 1367 (m), 1168 (m), 1114 (s), 996 (m), 859 (m), 822 (m), 704 (m); Anal. Calcd for C27H37NO6Si: C, 64.91; H, 7.44; N, 2.80. Found: C, 64.80; H, 7.43; N, 2.86.

1,6-Anhydro-4-deoxy-3,4-isopropylidene-4-t-butylcarbamide- β -D-altropyranose (5b).

To a solution of 4 (2.4 g, 5.0 mmol) in benzene (30.0 ml), 2,2-dimethoxypropane (20.0 ml, 162.7 mmol) and *p*-toluenesulfonic acid (0.1 g) as a catalyst were added, and the mixture was refluxed for 3 h. The reaction mixture was poured into saturated aqueous sodium bicarbonate and extracted with dichloromethane three times. The organic layer was dried over anhydrous magnesium sulfate, and the solvent was removed under reduced pressure. The residue was solved in 30.0 ml of tetrahydrofuran (THF). Further, 1.0 M tetrabutylammonium fluoride/THF (6.0 ml, 6.0 mmol) was added and the mixture was stirred at room temperature for 30 min. The reaction mixture was poured into saturated aqueous sodium bicarbonate and extracted with dichloromethane three times. The organic layer was dried over anhydrous magnesium sulfate, and the solvent was stirred at room temperature for 30 min. The reaction mixture was poured into saturated aqueous sodium bicarbonate and extracted with dichloromethane three times. The organic layer was dried over anhydrous magnesium sulfate, and the solvent was removed under reduced pressure. The residue was poured into saturated aqueous sodium bicarbonate and extracted with dichloromethane three times. The organic layer was dried over anhydrous magnesium sulfate, and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (*n*-hexane:ethyl acetate=2:1) to give **5b** (0.94 g, 62.7 % yield); mp 103-105°C; [α]²⁵_D -218.9° (c 0.52, CHCl3); ¹H-nmr (CDCl3) δ : 5.53 (1H, d, J=3.8 Hz, H-1), 4.92 (1H, m, H-5), 4.16 (1H, dd, J=7.3, 1.9)

Hz, H-3), 3.97-3.71 (4H, m, H-2,4,6), 2.48 (1H, d, *J*=9.2 Hz, O-H), 1.72-1.41 (6H, m, Me), 1.49 (9H, s, *t*-Bu); ir (KBr) 3460 (br), 2978 (m), 1694 (s), 1394 (s), 1369 (s), 1249 (m), 1158 (s), 1077 (s), 977 (m), 890 (m), 774 (m); Anal. Calcd for C14H23NO6: C, 55.81; H, 7.67; N, 4.64. Found: C, 55.64; H, 7.54; N, 4.75.

1,6-Anhydro-4-deoxy-3,4-isopropylidene-4-t-butylcarbamide-β-D-glycero-hexopyrano-2-ulose (6).

A solution of dimethyl sulfoxide (0.64 g, 8.2 mmol) in dry dichloromethane (4.0 ml) was slowly added dropwise to a solution of oxalyl chloride (0.46 g, 3.6 mmol) in dry dichloromethane (8.0 ml) at -60°C and the mixture was stirred at the same temperature for 15 min. Further, **5b** (0.45 g, 1.5 mmol) in dry dichloromethane (8.0 ml) was added dropwise to this mixture, stirring of the reaction mixture was continued for 40 min. After treatment of the mixture with triethylamine (2.3 g, 22.8 mmol), the reaction temperature was raised to 25°C. The reaction mixture was poured into water and dichloromethane, and extracted with dichloromethane. The organic layer was dried over anhydrous magnesium sulfate, and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (n-hexane:ethyl acetate=3:1) to give **6** (0.40 g, 88.3 % yield); mp 159-160.5°C; $[\alpha]^{25}$ D -125.5° (c 0.51, CHCl3); ¹H-nmr (CDCl3) δ : 5.28 (1H, s, H-1), 5.50 (1H, br s, H-5), 4.58 (1H, d, *J*=7.8 Hz, H-3), 4.30 (1H, d, *J*=7.8 Hz, H-4), 3.93 (1H, s, H-6), 3.91 (1H, s, H-6), 1.72-1.40 (6H, m, Me), 1.53 (9H, s, *t*-Bu); ir (KBr) 2976 (m), 1746 (s), 1694 (s), 1396 (m), 1371 (s), 1238 (m), 1174 (m), 1112 (m), 1093 (m), 969 (s), 872 (m), 808 (m), 756 (m); Anal. Calcd for C14H21NO6: C, 56.21; H, 7.05; N, 4.68. Found: C, 55.97; H, 6.93; N, 4.70.

3-Deoxy-2,3-isopropylidene-3-t-butylcarbamide-D-ribofuranose (8).

A solution of monoperoxyphthalic acid magnesium salt hexahydrate (0.59 g, 1.2 mmol) in methanol (4.0 ml) was added to 6 (0.49 g, 1.6 mmol) in dichloromethane (2.0 ml) at 0°C. The reaction temperature was raised to 25°C and the mixture was stirred overnight. After removing the precipitate by filtration, the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (*n*-hexane:ethyl acetate=1:1) to give a mixture of 7a and 7b (0.25 g). Under an argon atmosphere, 1.5 M diisobutylaluminium hydride/THF solution (3.5 ml, 5.3 mmol) was slowly added dropwise to this mixture (0.25 g) in dry THF (35.0 ml) at -78°C and the reaction solution was stirred at the same temperature for 1 h. A small amount of water was added and the temperature was raised to 25°C. Further, a large amount of THF was added and after drying the mixture over anhydrous magnesium sulfate, the precipitate was extracted with

THF and chloroform several times. The filtrate and extract were combined and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (*n*-hexane:ethyl acetate=1:1) to give **8** (0.15 g, 31.7 % yield) as a colorless oil; $[\alpha]^{25}D$ -52.7° (c 0.27, CHCl₃); ¹H-nmr (CDCl₃) δ : 5.44, 5.40 (0.5H, s, H-1a, b), 4.58-4.45 (2H, m, H-2,4), 4.33 (1H, m, H-3), 3.96-3.71 (2H, m, H-5), 1.69 (1.5H, s, Me), 1.63 (1.5H, s, Me), 1.59 (1.5H, s, Me), 1.48 (9H, s, *t*-Bu), 1.45 (1.5H, s, Me); ir (film) 3396 (br), 2980 (m), 2940 (m), 1707 (s), 1688 (s), 1396 (s), 1369 (s), 1257 (m), 1160 (m), 1077 (s); Anal. Calcd for C₁₃H₂₃NO₆: C, 53.98; H, 7.99; N, 4.84. Found: C, 53.92; H, 7.73; N, 4.86.

3-Amino-3-deoxy-1,2,3,5-tetraacetyl-D-ribofuranose (9).

Sulfuric acid (25 µl) was added to a solution of **8** (100 mg, 0.35 mmol) in acetic acid (3.0 ml) and acetic anhydride (1.3 ml, 13.8 mmol) at 0°C. After raising the temperature from 0 to 25°C, the reaction was continued overnight. The mixture was poured into saturated aqueous sodium bicarbonate (in the case of pH <7, addition of sodium bicarbonate is necessary) and extracted with chloroform three times. The solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (ethyl acetate) to give **9** (89 mg, 81.1 % yield) as a colorless oil; $[\alpha]^{25}D$ 46.1° (c 0.59 CHCl3); ¹H-nmr (CDCl3) δ : 6.44 (0.22H, d, *J*=4.3 Hz, H-1a), 6.12 (0.78H, s, H-b), 5.96 (0.22H, d, *J*=7.7 Hz, N-Ha), 5.65 (0.78H, d, *J*=8.9 Hz, N-Hb), 5.26 (0.22H, dd, *J*=7.7, 4.3 Hz, H-2a), 5.08 (0.78H, d, *J*=4.9 Hz, H-2b), 4.89 (0.78H, ddd, *J*=8.9, 8.9, 4.6 Hz, H-3b), 4.63 (0.22H, dd, *J*=7.9, 7.9, 3.9 Hz, H-3a), 4.11-4.05 (3H, m, H-4.5), 2.24-1.98 (12H, m, COCH₃); ir (film) 1746 (s), 1663 (m), 1543 (m), 1375 (m), 1224 (s), 1027 (m); hrms Calcd for C₁₃H₂₀NO8 (M⁺+1): 318.11889. Found: 318.122103.

9-(3'-Amino-3'-deoxy-2',3',5'-triacetyl-D-ribofuranosyl)-N⁶-benzoyladenine (10).

Under an argon atmosphere, 1,1,1,3,3,3-hexamethyldisilazane (8.5 ml, 40.3 mmol)and dry pyridine (1.9 ml, 23.5 mmol) were added to N^6 -benzoyladenine (263 mg, 1.10 mmol) and the mixture was refluxed for 5 h. The mixture was cooled and concentrated *in vacuo* for 3 h. A solution of **9** (225 mg, 0.71 mmol) in dry 1,2-dichloroethane (25.0 ml) was added to the residue at 25°C. Further, 1M SnCl4/dichloromethane (1.8 ml, 1.80 mmol) was slowly added dropwise, and the mixture was stirred at 80°C overnight. The mixture was poured into saturated aqueous sodium bicarbonate and extracted with chloroform-methanol (10:1) three times. The organic layer was dried over anhydrous magnesium sulfate and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (chloroform:methanol=15:1) and

NH-silica gel (chloroform:methanol=20:1) to give **10** (194 mg, 55.1 % yield) as a white solid; $[\alpha]^{25}D^{-4.7^{\circ}}$ (c 0.21 MeOH); ¹H-nmr (CDCl₃) δ : 8.98 (1H, s, N-H), 8.85 (1H, s, H-8), 8.16 (1H, s, H-2), 8.03 (2H, d, J=7.0 Hz, Ar-H), 7.63-7.50 (3H, m, Ar-H), 6.11 (1H, d, J=1.7 Hz, H-1'), 5.76 (1H, dd, J=6.1, 1.8 Hz, H-2'), 5.60 (1H, d, J=8.8 Hz, N-H), 5.42 (1H, m, H-3'), 4.55 (1H, d, J=10.1 Hz, H-5'), 4.37-4.23 (2H, m, H-4',5'), 2.23 (3H, s, COCH₃), 2.08 (6H, s, COCH₃), hrms Calcd for C_{23H25N6O7} (M⁺+1): 497.17847. Found: 497.18155.

3'-Amino-3'-deoxyadenosine (2).

A methanolic solution (1.6 ml) of **10** (20.2 mg, 0.04 mmol) containing 4.1 N methanolic NaOMe solution (0.3 ml) was refluxed for 15 h. The mixture was purified by column chromatography on iatrobeads (chloroform:methanol:25 % aq. NH3=30:10:1) to give **2** (9.2 mg, 85.2 % yield); mp 265-268°C (decomp.); $[\alpha]^{25}D$ -32.9° (c 0.24, 0.1N HCl) [lit.,¹⁵ mp 264-266°C (decomp.), $[\alpha]^{20}D$ -37°(0.1N HCl), lit.,¹⁶ mp 270-272°C (decomp.), $[\alpha]^{25}D$ -35° (0.1N HCl)]; ¹H-nmr (D₂O) δ : 8.21 (1H, s, H-8), 8.10 (1H, s, H-2), 5.98 (1H, d, *J*=3.2 Hz, H-1'), 4.51 (1H, dd, *J*=5.6, 3.2 Hz, H-2'), 3.97 (1H, m, H-4'), 3.84 (1H, dd, *J*=12.9, 2.3 Hz, H-5'), 3.69 (1H, dd, *J*=13.0, 2.3 Hz, H-5'), 3.52 (1H, dd, *J*=7.2, 5.6 Hz, H-3'), hrms Calcd for C₁₂H₁₅N₆O₃ (M⁺+1): 267.12056. Found: 267.12359.

Table 1. Summary of Crystal Data and Data Collection Parameters

orthorhombic P212121
0.20 x 0.20 x 0.15
a=11.2398 Å
b=25.5606 A
c=9.5142 A
V=2733.3921 Å ³
4
1.214 g cm ⁻³
MXCI ⁸
graphite
Ču Ka
w / 2q
128.7°
total: 2746
unique: 2646
0.037
0.043

X-Ray Diffraction of 4.

The crystal was mounted on a glass fiber. The cell constants and an orientation matrix for data collection, obtained from a least-squares refinement using a setting angle of 2 carefully centered reflections. Intensities was collected on an MXC1 diffractometer at room temperature. The data was corrected for lorentz and polarization effects. The crystal data and data collection parameters are summarized in Table 1. The structure was solved by the direct method using MITHRIL84 and expanded by Fourier Techniques using DIRDIF92. Full-matrix least-squares refinement included anisotropic thermal parameters for non-H atoms as well as isotropic thermal parameters for H atoms. Convergence was reached at R=0.037 and $R_w=0.043$.

ACKNOWLEDGMENTS

We would like to thank Mr. Masayoshi Kusama for his mass spectroscopic analysis. We are also grateful to Mr. Hideo Ago for the X-ray analysis.

REFERENCES

- 1 This compound is avaible from Yuki Gousei Kogyo Co., Ltd.
- ² 'Levoglucosenone and Levoglucosans, Chemistry and Applications', ed. by Z. J. Witczak, ATL
 Press, Inc., USA, 1994, pp.3-118, and references cited therein.
- a) K. Koseki, T. Ebata, H. Kawakami, H. Matsushita, Y. Naoi, and K. Itoh, <u>Heterocycles</u>, 1990, 31, 423, b) T. Ebata, K. Matsumoto, H. Yoshikoshi, K. Koseki, H. Kawakami, and H. Matsushita, <u>Heterocycles</u>, 1990, 31, 1585, c) K. Matsumoto, T. Ebata, K. Koseki, and H. Matsushita, <u>Bull. Chem.</u> <u>Soc. Jpn.</u>, 1991, 61, 2309, d) K. Matsumoto, T. Ebata, K. Koseki, and H. Matsushita, <u>Heterocycles</u>, 1991, 32, 2225, e) K. Okano, T. Ebata, K. Koseki, H. Kawakami, K. Matsumoto, and H. Matsushita, <u>Chem. Pharm. Bull</u>., 1993, 41, 861, f) K. Matsumoto, T. Ebata, and H. Matsushita, <u>Carbohydr. Res.</u>, in press.
- 4 N. N. Gerber and H. A. Lechevalier, <u>J. Org. Chem.</u>, 1962, **27**, 1731.
- 5 A. J. Guarrino and N. M. Kredich, <u>Biochim. Biophys. Acta</u>, 1963, 68, 317.
- 6 B. M. Cassy and R. J. Suhadolnik, <u>Biochim. Biophys. Acta</u>, 1969, **182**, 316.
- 7 R. J. Suhadolnik and B. M. Cassy, Biochim. Biophys. Acta, 1969, 179, 258.

- a) R. J. Suhadolnik, 'Nucleoside Antibiotics,' Wiley-Interscience: New York, 1970: b) R. J.
 Suhadolnik, 'Nucleoside as Biological Probes,' Wiley-Interscience: New York, 1979.
- J. T. Trauman and H. Klenow, Mol. Pharmacol., 1968, 4, 77.
- 10 V. W. Armstrong and F. Eckstain, Biochemistry, 1979, 18, 5117.
- 11 T. Kutateladze, R. Beabealashvili, A. Azhayev, and A. Krayevsky, FEBS Lett., 1983, 153, 420.
- a) B. R. Baker, R. E. Schaub, and J. H. Williams, <u>J. Am. Chem. Soc</u>., 1955, 77, 7; b) B. R. Baker, R. E.
 Schaub, and H. M. Kissman, <u>J. Am. Chem. Soc</u>., 1955, 77, 5911.
- 13 R. Mengel and H. Wiedner, <u>Chem. Ber.</u>, 1976, **109**, 433.
- 14 M. Ikehara, T. Maruyama, and H. Miki, <u>Tetrahedron Lett.</u>, 1976, 4485.
- 15 R. Ranganathan and D. Larwood, <u>Tetrahedron Lett.</u>, 1978, 4341.
- 16 M. C. Samano and M. J. Robinss, Tertahedron Lett., 1989, 30, 2329.
- M. J. Robins, S. D. Hawrelak, A. E. Hernandez, and S. F. Wnuk, <u>Nucleosides & Nucleotides</u>, 1992, 11, 821.
- 18 F. W. Lichtenthaler, E. Cuny, T. Morino, and I. Rychlik, <u>Chem. Ber.</u>, 1979, **112**, 2588.
- 19 A. Okruszek and G. Verkade, <u>J. Med. Chem.</u>, 1979, **22**, 882.
- 20 E. Herranz, S. A. Biller, and K. B. Sharpless, J. Am. Chem. Soc., 1978, 100, 3596.
- 21 A. J. Mancuso, S. -L. Huang, and D. Swern, J. Org. Chem., 1978, 43, 2480.

Received, 28th November, 1994