PARATOCARPINS F - L, SEVEN NEW ISOPRENOID-SUBSTITUTED FLAVONOIDS FROM *PARATOCARPUS VENENOSA* ZOLL¹

Yoshio Hano, Naoyuki Itoh, Akio Hanaoka, and Taro Nomura* Faculty of Pharmaceutical Sciences, Toho University, 2-2-1, Miyama, Funabashi, Chiba 274, Japan

Abstract — Two new isoprenoid substituted chalcones, paratocarpins F (1) and G (2), along with five new isoprenoid substituted flavanones, paratocarpins H (3), I (4), J (5), K (6), and L (7), were isolated from the Indonesian moraceous plant, *Paratocarpus* (= *Artocarpus*) venenosa Zoll. The structures of paratocarpins F, G, H, I, J, K, and L were shown to be 1, 2, 3, 4, 5, 6, and 7, respectively, on the basis of spectroscopic and chemical evidence.

Previously we reported the structure determination of five isoprenoid-substituted chalcones, paratocarpins A - E, isolated from *Paratocarpus* (= *Artocarpus*) venenosa Zoll.² Further extension of studies on the components of *P. venenosa* led to the isolation of two new isoprenoid-substituted chalcones, paratocarpins F (1) and G (2), along with five new isoprenoid-substituted flavanones, paratocarpins H (3), I (4), J (5), K (6), and L (7). This paper deals with characterization of these new flavonoids as well as the known compounds, gancaonin Q³ (8) and 6-prenylapigenin (9).⁴

Paratocarpin F (1), yellow prisms, mp 89 - 91 °C, C25H26O5, showed positive reaction to methanolic ferric chloride reaction. The ir spectrum disclosed absorption bands due to hydroxyl, conjugated carbonyl, and benzene ring moieties. The uv spectrum of 1 exhibited maxima at 210, 225 (sh), 289, and 377 nm, and was similar to that of paratocarpin C² (10), indicating that 1 is a 2', 4, 4'-oxygenated chalcone derivative. The ¹H nmr spectrum of 1 showed the signals of the following protons (acetone-*d*6, 400 MHz): protons in a 2,2-dimethylpyran ring, δ 1.45 (6H, s), 5.83, 6.47 (each 1H, d, J = 10 Hz), orthocoupled aromatic protons, δ 6.40, 8.09 (each 1H, d, J = 9 Hz), ABX type aromatic protons, δ 6.81 (1H, d, J = 8 Hz), 7.60 (1H, d, J = 2 Hz), 7.64 (1H, dd, J = 2 and 8 Hz), two olefinic protons, δ 7.81 (2H, s), proton in a hydrogen-bonded hydroxyl group, δ 13.66 (1H, s), protons in a 2-(1-hydroxy-1-methylethyl)-



Table 1 ¹³C Nmr chemical shifts (ppm) of 1, 2, 10, 11, and 12 (δ in acetone-d₆)

C	1	2	10	11	12
C-1	128.8	128.8	128.8	127.6	127.7
C-2	127.8	127.0	127.8	133.6	131.7
C-3	122.4	132.5	122.4	127.7	129.7
C-4	156.5	163.7	156.4	159.9	158.7
C-5	117.5	111.0	117.5	117.5	116.4
C-6	131.4	132.6	131.3	130.1	129.2
C-α	119.2	118.7	119.2	118.3	118.2
C-β	144.8	145.0	144.6	145.2	145.4
C=0	193.0	193.0	193.0	193.1	193.1
C-1'	115.5	114.5	114.5	114.5	114.4
C-2'	168.3	165.2	165.2	165.2	165.2
C-3'	114.7	116.2	116.2	116.2	116.2
C-4'	162.5	162.8	162.8	162.7	162.7
C-51	102.4	108.1	108.1	108.0	108.0
C-61	133.2	130.3	130.3	130.2	130.2
C-1"	27.5	22.3	22.3	22.3	22.3
C-2"	92.6	123.3	123.3	123.3	123.3
C-3"	71.5	131.5	131.5	131.5	131.5
C-4"	25.5	17.9	17.9	17.9	17.9
C-5"	26.0	25.9	25.9	25.9	25.9
C-6"	122.3	72.9	122.4	38.7	29.1
C-7"	132.5	98.9	132.4	76.7	123.3
C-8"	78.0	71.1	78.0	148.4	132.8
C-9"	28.4	25.8	28.4	110.8	17.9
C-10 "	28.4	26.4	28.4	18.3	25.9

Assignments were performed by 2D 13 C- 1 H shift correlation spectroscopic methods (CHCOSY, COLOC, HMBC).



Figure 2 NOESY spectrum of 1

2,3-dihydrofuran ring,⁵ δ 1.24, 1.29 (each 3H, s), 3.12 (1H, dd, J = 10 and 16 Hz), 3.19 (1H, dd, J = 8 and 16 Hz), 4.81 (1H, dd, J = 8 and 10 Hz). The ¹³C nmr spectrum of 1 was analysed by comparing with that of 10 (Table 1). In the spectrum, the chemical shifts of all the carbon atoms except those of the carbons of the A ring and isoprenoid moiety on the ring were good agreement with those of the relevant carbons of 10. This result supports that the 2-(1-hydroxy-1-methylethyl)-2,3-dihydrofuran ring locates on the A ring. Furthermore the location of the 2,2-dimethylpyran ring was confirmed by the two dimetional NOE spectroscopy (NOESY) spectrum of 1 as described in Figure 2. From above results, the structure of paratocarpin F is characterized as 1.

Paratocarpin G (2), yellow needles, mp 113 - 116 °C, C₂₅H₂₈O₆, showed positive reaction to methanolic ferric chloride reaction. The uv spectrum of **2** exhibited maxima at 204, 240 (sh), 315 (sh), 369 nm, and was similar to those of paratocarpin D (11)² and kanzonol C (12).^{6,7} The ¹H nmr spectrum of **2** showed the signals of the following protons (acetone-*d*₆, 400 MHz): protons in a 3,3-dimethylallyl group, δ 1.65, 1.78 (each 3H, s), 3.38 (2H, br d, J = 7 Hz), 5.28 (1H, m), *ortho*-coupled aromatic protons, δ 6.54, 8.00 (each 1H, d, J = 9 Hz), ABX type aromatic protons, δ 6.86 (1H, d, J = 8 Hz), 7.71 (1H, dd, J = 2 and 8 Hz), 7.88 (1H, d J = 2 Hz), two olefinic protons, δ 7.79, 7.87 (each 1H, d, J = 15 Hz), proton in a hydrogen-bonded hydroxyl group, δ 13.97 (1H, s), two methyl protons, δ 1.27, 1.29 (each 3H, s), two methine protons, δ 4.36 (1H, d, J = 4 Hz), 5.45 (1H, dd, J = 4 and 7 Hz), proton in a hydroxyl group, δ 4.84 (1H, d, J = 7 Hz). The ¹³C nmr spectrum of **2** was analysed by comparing with those of **11** and **12** as described in Table 1. In the spectrum of **2**, the chemical shifts of all the carbon atoms except those of the carbons of **12**. The presence of 2-(1-hydroxy-1-methylethyl)-3-hydroxy-2,3-dihydrofuran ring in the structure along with the locations of the ring and the 3,3-dimethylallyl group was confirmed by the HMBC spectrum as described in Figure 3. The relative configuration between 6"-H and 7"-H was



Figure 3 HMBC spectrum of 2 ($J_{CCH} = 6$ Hz)



Figure 4 NOESY spectrum of 2

supported by consideration of the NOESY spectrum of 2 (Figure 4). In the spectrum, the NOE was observed between the 6"-H and the 9"-H as well as the 10"-H. This result supports the relative configuration between 6"-H and 7"-H to be *trans*. From the above results, the structure of paratocarpin G was represented by the formula (2).

Paratocarpin H (3), colorless prisms, mp 211 - 212 °C, C25H26O5, showed positive reaction to methanolic ferric chloride reaction, magnesium-hydrochloric acid test, and sodium borohydride test.⁸ The uv spectrum exhibited maxima at 225, 294, 340 (sh) nm, and was similar to that of naringenin (13).⁹ The ¹H nmr spectrum of 3 showed the signals of the following protons (acetone-*d*6, 400 MHz): protons in a 3,3-dimethylallyl group, δ 1.64, 1.75 (each 3H, s), 3.25 (2H, br d, J = 7 Hz), 5.24 (1H, m), protons in a 2,2-dimethylypran ring, δ 1.42 (6H, s), 5.77, 6.44 (each 1H, d, J = 10 Hz), ABX type protons, δ 2.73 (1H, dd, J = 3 and 17 Hz), 3.16 (1H, dd, J = 13 and 17 Hz), 5.41 (1H, dd, J = 3 and 13 Hz), an aromatic proton, δ 6.04 (1H, s), ABX type aromatic protons, δ 6.78 (1H, d, J = 8 Hz), 7.23 (1H, d, J = 2 Hz), 7.29 (1H, dd, J = 2 and 8 Hz), proton in a hydrogen-bonded hydroxyl group, δ 12.46 (1H, s). The ¹³C





















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Figure 5

С	3 ^a	4 ^b	5 ^a	6 ^b	7 ^a
C-2	79.7	79.1	79.9	78.9	80.1
C-3	43.6	43.2	43.7	43.2	43.7
C-4	197.2	196.1	197.6	196.0	197.4
C-4a	103.3	102.9	103.0	102.9	103.2
C-5	162.3	162.5	162.2	162.4	162.3
C-6	109.1	103.0	102.6	103.1	109.4
C-7	164.8	158.4	161.7	158.5	164.8
C-8	95.3	96.3	96.6	96.3	95.4
C-8a	162.0	162.1	163.6	162.2	162.1
C-1'	132.4	130.5	131.0	130.7	131.0
C-2'	125.7	128.2	129.0	128.0	129.1
C-3'	122.2	127.3	121.9	115.7	129.0
C-4'	154.2	154.9	155.4	156.1	156.2
C-5'	117.0	116.0	118.0	115.7	115.8
C-6'	128.4	125.7	126.6	128.0	126.2
C-1"	21.6	115.3	16.3	115.3	21.7
C-2"	123.6	126.2	32.4	126.3	123.5
C-3"	131.3	78.3	75.1	78.3	131.2
C-4"	25.9	28.4	26.8	28.4	25.9
C-5"	17.8	28.4	27.0	28.4	17.9
C-6"	122.7	29.9	23.1		29.1
C-7"	132.2	121.2	33.3		123.6
C-8"	77.2	135.4	77.0		132.7
C-9"	28.2	25.8	27.1		25.9
C-10"	28.2	17.9	27.1		17.9

Table 2 ¹³C Nmr chemical shifts (ppm) of 3, 4, 5, 6, and 7

Assignments were performed by 2D shift correlation spectroscopic methods (CHCOSY, COLOC, HMBC)

Solvent: a; acetone-d6 b; CDCl3



Figure 6 COLOC spectrum of 3 ($J_{CCH} = 6 \text{ Hz}$)

nmr spectrum of **3** was analysed by comparing with those of flavanone derivatives¹⁰ (Table 2). From above results, compound **3** is a 5,7,4'-oxygenated flavanone derivative. The location of the 3,3-dimethylallyl group and the 2,2-dimethylpyran ring was confirmed by the COLOC spectrum (Figure 6). From the above results, the structure of paratocarpin H is characterized as **3**.

Paratocarpin I (4), pale yellow oily substance, C25H26O5, showed positive reaction to methanolic ferric chloride reacion, magnesium-hydrochloric acid test, and sodium borohydride test. The uv spectrum exhibited maxima at 203, 226, 272, 294, 305 (sh), 360 nm, and was similar to euchrenone a_2 (14).¹¹ The ¹H nmr spectrum of 4 showed the signals of the following protons (acetone-*d*6, 400 MHz): protons in a 3,3-dimethylallyl group, δ 1.78 (6H, s), 3.38 (2H, br d, J = 7 Hz), 5.32 (1H, m), protons in a 2,2-dimethylpyran ring, δ 1.43, 1.44 (each 3H, s), 5.50, 6.62 (each 1H, d, J = 10 Hz), ABX type protons, δ 2.76 (1H, dd, J = 3 and 17 Hz), 3.08 (1H, dd, J = 13 and 17 Hz), 5.31 (1H, dd, J = 3 and 13 Hz), an aromatic proton, δ 5.95 (1H, s), ABX type aromatic protons, δ 6.84 (1H, d, J = 8 Hz), 7.18 (1H, d, J = 2 Hz), 7.19 (1H, dd, J = 2 and 8 Hz), proton in a hydrogen-bonded hydroxyl group, δ 12.31 (1H, s). Comparative examination of the EI-mass spectra of **3** and **4** was carried out. The EI-mass spectrum of **3** showed the fragment ion at *m*/z 205 (15), while that of **4** showed at *m*/z 203 (16). This result supports the 2,2-dimethylpyran ring to be on the A ring. The ¹³C nmr spectrum of **4** was analysed (Table 2). Fukai, *et al* reported that the chemical shift of the C-1 atom of the 3,3-dimethylallyl group was depend on the substituents located at the adjacent positions.¹² The signal of the C-1 atom of the 3,3-dimethylallyl group



Figure 7 HMBC spectrum of 4 ($J_{CCH} = 6$ Hz)

of 4 was observed at δ 29.9. This result suggests that one of the *ortho*-positions to the 3,3-dimethylallyl group is replaced by the oxygenated substituent, and the group locates at the C-3' position. The location of the 2,2-dimethylpyran ring was confirmed by the HMBC spectrum of 4 (Figure 7). Thus, the structure of paratocarpin I is characterized as 4.

Paratocarpin J (5), colorless oily substance, C25H28O5, showed positive reaction to methanolic ferric chloride reaction, magnesium-hydrochloric acid test, and sodium borohydride test. The uv spectrum exhibited maxima at 204, 213 (sh), 228 (sh), 294, 330 (sh) nm, and was similar to that of **13**. The ¹H nmr spectrum of **5** showed the signals of the following protons (acetone-*d*6, 400 MHz): protons in two sets of 2,2-dimethyldihydropyran ring, δ 1.32 (12H, s), 1.83, 1.84, 2.58, 2.85 (each 2H, t, J = 7 Hz), ABX type protons, δ 2.75 (1H, dd, J = 3 and 17 Hz), 3.18 (1H, dd, J = 13 and 17 Hz), 5.40 (1H, dd, J = 3 and 13 Hz), an aromatic proton, δ 5.85 (1H, s), ABX type aromatic protons, δ 6.76 (1H, d, J = 8 Hz), 7.23 - 7.25 (2H, m), a proton in a hydrogen-bonded hydroxyl group, δ 12.56 (1H, s). The EI-mass spectrum of **5** showed the fragment ion at m/z 220 (**17**) and 188 (**18**). The location of the 2,2-dimethyl-dihydropyran ring in the A ring was confirmed by the HMBC spectrum (Figure 8). Thus, the structure of



Figure 8 HMBC spectrum of 5 ($J_{CCH} = 6$ Hz)



Figure 9 Chemical correlation of compound (7) to compound (5)

paratocarpin J is characterized as 5. The structure (5) was confirmed by the derivation from paratocarpin L (7) (Figure 9). This compound (5) has been synthesized by Mitscher *et al.*¹³ To our knowledge, this is the first time that it has been identified as a natural product.

Paratocarpin K (6), pale yellow prisms, mp 165 - 166 °C, C20H18O5, showed positive reaction to methanolic ferric chloride, magnesium-hydrochloric acid test, and sodium borohydride test. The uv spectrum exhibited maxima at 227, 272, 295, 307, 360 nm, and was similar to those of 4 and citflavanone (19).¹⁴ The ¹H nmr spectrum of 6 showed the signals of the following protons (CDCl3, 400 MHz): protons in a 2,2-dimethylpyran ring, δ 1.43, 1.44 (each 3H, s), 5.50, 6.62 (each 1H, d, J = 10 Hz), ABX type protons, δ 2.78 (1H, dd, J = 3 and 17 Hz), 3.07 (1H, dd, J = 13 and 17 Hz), 5.34 (1H, dd, J = 3 and 13 Hz), A2B2 type aromatic protons, δ 6.88, 7.32 (each 2H, d, J = 8 Hz), an aromatic proton, δ 5.95 (1H, s), a proton in a hydrogen-bonded hydroxyl group, δ 12.29 (1H, s).. The EI-mass spectrum of 6 showed the fragment ion m/z 218 (20). The ¹³C nmr spectrum of 6 was analysed by comparing with that of 4 (Table 2). In the spectrum, the chemical shifts of all the carbon atoms except those of the B ring carbons were similar to those of the relevant carbons of 4. From the above results, the structure of paratocarpin K is characterized as 6. While the compound (6) has been synthesized by Jain *et al*, ¹⁵ this is the first time that it has been identified as a natural product.

Paratocarpin L (7), colorless needles, mp 169 - 170 °C, C₂₅H₂₈O5, gave a dark blue color with methanolic ferric chloride. The uv spectrum of **7** exhibited maxima at 207, 228 (sh), 292, 334 (sh) nm, indicating that **7** is a flavanone derivative.⁹ The ¹H nmr of **7** showed the signals of the following protons (acetone-*d*₆, 400 MHz): protons in two sets of 3,3-dimethylallyl groups, δ 1.64 (3H, s), 1.71 (6H, s), 1.75 (3H, s), 3.25, 3.35 (each 2H, br d, J = 7 Hz), 5.23, 5.35 (each 1H, m), ABX type protons, δ 2.70 (1H, dd, J = 3 and 17 Hz), 3.15 (1H, dd, J = 13 and 17 Hz), 5.39 (1H, dd, J = 3 and 13 Hz), an aromatic proton, δ 6.02 (1H, s), ABX type aromatic protons, δ 6.88 (1H, d, J = 8 Hz), 7.20 (1H, dd, J = 2 and 8 Hz), 7.28 (1H, d, J = 2 Hz), a proton in a hydrogen-bonded hydroxyl group, δ 12.47 (1H, s). The ¹³C nmr spectrum of **7** was analysed by comparing with those of flavanones, indicating that **7** is a 5,7,4'-trioxygenated flavanone derivative (Table 2). The EI-mass spectrum of **7** showed the fragment ion at *m/z* 220 and 165 (**21**). These results suggest that the structure of paratocarpin L is characterized as **7** or **7a**. Paratocarpin L was identified with **7** which was derived from 2-hydroxy-4,6-dimethoxymethoxy-5-prenylacetophenone (**22**)¹⁶ and 4-methoxymethoxy-3-prenylbenzaldehyde (**23**)¹⁷ (Figure 10). Thus, the structure of paratocarpin L was represented by formula(**7**).



Figure 10 Synthesis of compound (7)

EXPERIMENTAL

Abbreviations: s = singlet, d = doublet, dd = double doublet, m = multiplet, br = broad, sh = shoulder. The general procedures followed and the instruments used in our previous paper.²

Isolation of Paratocarpins F (1), G (2), H (3), I (4), J (5), K (6), and L (7)

The dried bark of *P. venenosa* (3.4 kg) was extracted at room temperature with *n*-hexane (9 1, three times), benzene (9 1 x 3), and acetone (9 1 x 3), successively (each 3 days). Evaporation of the *n*-hexane, benzene and acetone solutions to dryness yielded 60 g, 55 g, and 95 g of the residues, respectively.²

The *n*-hexane extract (30 g) was chromatographed over silica gel (300 g) using *n*-hexane, *n*-hexane - ethyl acetate (97 : 3, 95 : 5, 4 : 1), and then ethyl acetate to prepare frs. 1 - 46.² The fraction eluted with *n*-hexane - ethyl acetate (95 : 5, frs. 25 and 26, 18 mg) was fractionated by preparative tlc (benzene) to give paratocarpin J (5, 2 mg). The fraction eluted with ethyl acetate (frs. 41 - 46, 5 g) was rechromatographed over silica gel (150 g) with *n*-hexane containing increasing amount of ethyl acetate as an eluent (frs. 1' - 58'). The fraction eluted with *n*-hexane - ethyl acetate (4 : 1, frs. 20' - 24', 540 mg) was fractionated by preparative tlc [*n*-hexane - acetone (2 : 1)] to give paratocarpin L (7, 36 mg).

The benzene extract (25 g) was chromatographed over silica gel (300 g) using *n*-hexane, *n*-hexane - ethyl acetate (99 : 1, 98 : 2, 96 : 4, 95 : 5, 9 : 1, and 85 : 15) to prepare frs. 1" - 98".² The fraction eluted with *n*-hexane - ethyl acetate (98 : 2, frs. 19" and 20", 70 mg) was fractionated by preparative tlc (benzene) to give paratocarpin J (5, 2 mg). The fraction eluted with *n*-hexane - ethyl acetate (95 : 5, fr. 37", 150 mg) was fractionated by preparative tlc [*n*-hexane - ethyl acetate (5 : 1)] to give paratocarpin H (3, 10 mg). The fraction eluted with *n*-hexane - ethyl acetate (5 : 1)] to give paratocarpin I (4, 6 mg). The fraction eluted with *n*-hexane - ethyl acetate (9 : 1, frs. 60" - 65", 210 mg) was fractionated by preparative tlc [*n*-hexane - acetone (2 : 1)] to give paratocarpin F (1,

6 mg). The fraction eluted with *n*-hexane - ethyl acetate (9 : 1, frs. 66" - 69", 200 mg) was fractionated by preparative tlc [*n*-hexane - ethyl acetate (3 : 1)] to give paratocarpin K (6, 7 mg). The fraction eluted with *n*-hexane - ethyl acetate (85 : 5, frs. 74" and 75", 800 mg) was fractionated by preparative tlc [*n*-hexane - ethyl acetate (2 : 1)] to give paratocarpin L (7, 210 mg). The acetone extract (30 g) was chromatographed over silica gel (300 g) using benzene, benzene - acetone (95 : 5, 92 : 8, and 8 : 2) to prepare frs. 1"' - 55"'. The fraction eluted with benzene (fr. 6"', 210 mg) was fractionated by preparative tlc [*n*-hexane - acetone (3 : 1)] to give paratocarpins H (3, 5 mg) and I (4, 3 mg). The fraction eluted with benzene - acetone (95 : 5, fr. 12"', 95 mg) was fractionated by preparative tlc [*n*-hexane - acetone (2 : 1)] to give gancaonin Q³ (8, 9 mg). The fraction eluted with benzene - acetone (92 : 8, fr. 17"', 164 mg) was fractionated by preparative tlc [CHCl₃ - MeOH (50 : 1)] to give paratocarpin G (2, 26 mg). The known compound, gancaonin Q³ (8), was identified by direct comparison with the authentic sample. 6-Prenylapigenin (9) was identified by comparison with the published data⁴ and the authentic sample (9a) which was derived from naringenin¹⁸ (13).

Paratocarpin F (1)

Compound (1) was recrystallized from MeOH - H₂O (2 : 1) to give yellow prisms, mp 89 - 91 °C. FeCl₃ test: positive (dark brown). $[\alpha]_{D}^{22}$ -6.9° (c = 0.1, MeOH). Uv λ_{max}^{MeOH} nm (log ε): 210 (4.53), 225 (sh 4.42), 289 (4.20), 377 (4.59). EI- ms: m/z (rel. int.) 406 (M⁺, 24), 391 (100), 373 (3), 347 (4), 338 (8), 203 (6), 187 (6), 171 (31). HR-ms: m/z 406.1759 (M⁺, C₂₅H₂₆O₅ requires 406.1780), 391.1516 (C₂₄H₂₃O₅ requires 391.1546). Ir ν_{max}^{KBr} cm⁻¹: 3430, 2975, 2930, 1640, 1600, 1560, 1490, 1430.

Paratocarpin G (2)

Compound (2) was recrystallized from MeOH to give yellow needles, mp 113 - 116 °C. FeCl₃ test: positive (dark brown). $[\alpha]_{D}^{2}2^{0^{\circ}}(c = 0.1, \text{ MeOH})$. Uv $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 204 (4.66), 240 (sh 4.23), 315 (sh 4.23), 369 (4.62). EI- ms: m/z (rel. int.) 424 (M⁺, 37), 407 (2), 406 (2), 381 (11), 369 (4), 348 (19), 333 (7), 305 (67), 293 (23), 204 (23), 203 (23), 189 (18), 187 (28), 161 (42), 149 (100). HR-ms: m/z 424.1855 (M⁺, C₂₅H₂₈O₆ requires 424.1886), 305.0796 (C₁₉H₁₃O₄ requires 305.0814). Ir $\nu_{\text{MB}}^{\text{MB}}$ cm⁻¹: 3430, 2930, 1630, 1610, 1560, 1490, 1440.

Paratocarpin H (3)

Compound (3) was recrystallized from *n*-hexane - CHCl₃ (1 : 1) to give colorless prisms, mp 211 - 212 °C. FeCl₃ test: positive (dark brown). Mg - HCl test: positive (violet). NaBH₄ test: positive (orange). $[\alpha]_D^{22}$ -26.1° (c = 0.1, MeOH). Uv λ_{max}^{MeOH} nm (log ϵ): 225 (4.57), 294 (4.14), 340 (sh 3.39). EI- ms: m/z (rel. int.) 406 (M⁺, 37), 391 (93), 363 (6), 351 (9), 205 (20), 171 (100). HR-ms: m/z 406.1781 (M⁺, C₂₅H₂₆O₅ requires 406.1780), 391.1516 (C₂₄H₂₃O₅ requires 391.1546). Ir ν_{max}^{KBr} cm⁻¹: 3400 (br), 3140, 2970, 2920, 1650, 1630, 1590, 1490, 1450.

Paratocarpin I (4)

Compound (4) was obtained as oily substance. FeCl₃ test: positive (dark brown). Mg - HCl test: positive (violet). NaBH₄ test: positive (orange). $[\alpha]_D^{22}$ 0° (c = 0.1, MeOH). Uv λ_{max}^{MeOH} nm (log ε): 203 (4.63), 226 (4.26), 272 (4.48), 294 (4.00), 305 (sh 3.89), 360 (3.50). El-ms: m/z (rel. int.) 406 (M⁺, 22), 391 (79), 375 (34), 293 (3), 203 (100), 187 (67). HR-ms: m/z 406.1771 (M⁺, C₂₅H₂₆O₅ requires 406.1780), 391.1542 (C₂₄H₂₃O₅ requires 391.1546). Ir v_{max}^{KBr} cm⁻¹: 3430 (br), 2930, 1650, 1570, 1500, 1450.

Paratocarpin J (5)

Compound (5) was obtained as oily substance. FeCl₃ test: positive (dark brown). Mg - HCl test: positive (violet). NaBH₄ test: positive (orange). $[\alpha]_D^{22}0^\circ$ (c = 0.1, MeOH). Uv $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 204 (4.55), 213 (sh 4.31), 228 (sh 4.25), 294 (4.13), 330 (sh 3.21). EI-ms: m/z (rel. int.) 408 (M⁺, 37), 353 (3), 220 (12), 205 (6), 188 (62), 175 (100), 165 (37), 149 (9). HR-ms: m/z 408.1949 (M⁺, C₂₅H₂₈O₅ requires 408.1937). Ir v_{max}^{KBr}cm⁻¹: 3440 (br), 1640, 1580, 1500, 1480, 1440.

Formation of 5 from 7

A mixture of 7 (50 mg) and 35% HCl (3 ml) - MeOH (15 ml) solution was refluxed for 2 h, and treated as usual. The product was purified by preparative tlc (benzene) to give 5 (15.5 mg). The compound (5) was identified with 5 by comparing the phisical data of 5 with those of 5.

Paratocarpin K (6)

Compound (6) was recrystallized from *n*-hexane - CHCl₃ (1 : 1) to give yellow proisms, mp 165 - 166 °C. FeCl₃ test: positive (dark brown). Mg - HCl test: positive (violet). NaBH₄ test: positive (orange). $[\alpha]_D^{22}$ 0° (c = 0.1, MeOH). Uv λ_{max}^{MeOH} nm (log ϵ): 227 (4.15), 272 (4.51), 295 (3.97), 307 (3.87), 360 (3.33). EI- ms: *m/z* (rel. int.) 338 (M⁺, 13), 323 (49), 218 (3), 203 (100), 147 (3), 135 (2), 120 (5) HR-ms: *m/z* 338.1131 (M⁺, C₂₀H₁₈O₅ requires 338.1155), 203.0354 (C₁₁H₇O₄ requires 203.0344). Ir ν_{max}^{KBr} cm⁻¹: 3430 (br), 2930, 1650, 1570, 1520, 1450.

Paratocarpin L (7)

Compound (7) was recrystallized from *n*-hexane - ethyl ether (1 : 1) to give pale yellow needles, mp 169 - 170 °C. FeCl₃ test: positive (dark brown). Mg - HCl test: positive (violet). NaBH₄ test: positive (orange). $[\alpha]_D^{22}$ -11° (*c* = 0.04, EtOH). Uv $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 205 (4.35), 210 (4.31), 228 (4.14), 295 (3.99), 336 (3.27). EI- *ms*: *m/z* (rel. int.) 408 (M⁺, 100), 393 (3), 365 (18), 353 (27), 220 (25), 205 (45), 192 (25), 175 (19), 165 (56). HR-ms: *m/z* 408.1936 (M⁺, C₂₅H₂₈O₅ requires 408.1929), 165.0175 (C₈H₅O₄ requires 165.0186). Ir $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450, 3150, 1640, 1600, 1500.

Formation of 24 from 2-hydroxy-4,6-dimethoxymethoxy-5-prenylacetophenone (22) and 4-methoxymethoxy-3-prenylbenzaldehyde

A mixture of 22^{16} (110 mg) and 23^{17} (95 mg) in EtOH solution (3 ml) and 25% KOH aqueous solution (1 ml) was kept at room temperature for 15h, and treated as usual. The product was purified by preparative tlc [*n*-hexane - acetone (5 : 1)] to give 24 (21 mg). Compound (24) was obtained as yellow oily substance. HR-ms: m/z 540.2687 (M⁺, C₃₁H₄₀O₈ requires 540.2712). ¹H Nmr (acetone- d_6 , 400 MHz): δ 1.65, 1.75, 1.76, 1.81 (each 3H, s), 3.39 (4H, br d, J = 7 Hz), 3.47 (6H, s, -OCH₂OCH₃ x 2), 3.50 (3H, s, -OCH₂OCH₃), 4.96 (2H, s, -OCH₂OCH₃), 5.33 (4H, s, -OCH₂OCH₃ x 2), 5.23, 5.35 (each 1H, m), 6.48 (1H, s), 7.16 (1H, d, J = 8 Hz), 7.58 (1H, dd, J = 2 and 8 Hz), 7.59 (1H, d, J = 2 Hz), 7.80 (2H,s), 12.99 (1H, s).

Formation of 25 from 24.

A mixture of 24 (17 mg) and sodium acetate (42 mg) in EtOH (5 ml) was refluxed for 2h, and treated as usual. The product was purified by preparative tic [*n*-hexane - acetone (7 : 1)] to give 25 (5 mg). Compound (25) was obtained as yellow oily substance. HR-ms: m/z 540.2699 (M⁺, C₃₁H₄₀O₈ requires 540.2712). Ir v_{max}^{KBr} cm⁻¹: 1670, 1600, 1160. ¹H Nmr (acetoned₆, 400 MHz): δ 1.65, 1.71, 1.73, 1.78 (each 3H, s), 2.67 (1H, dd, J = 3 and 17 Hz), 3.04 (1H, dd, J = 13 and 17 Hz), 3.38,

2325

3.39 (each 2H, br d, J = 7 Hz), 3.45 (6H, s, $-OCH_2OCH_3 \times 2$), 3.53 (2H, s, $-OCH_2OCH_3$), 5.01, 5.14 (each 1H, d, J = 7 Hz, $-OCH_2OCH_3$), 5.20 (1H, m), 5.27, 5.31 (each 2H, s, $-OCH_2OCH_3$), 5.32 (1H, m), 5.47 (1H, dd, J = 3 and 13 Hz), 6.51 (1H, s), 7.12 (1H, d, J = 8 Hz), 7.34 (1H, dd, J = 2 and 8 Hz), 7.35 (1H, d, J = 2 Hz).

Formation of 7 from 25

A mixture of 25 (10 mg) and 3N HCl solution (0.2 ml) was refluxed for 30 min, and treated as usual. The product was purified by preparative tlc [*n*-hexane - acetone (2 : 1)] to give 7 (3 mg). The compound (7) was recrystallized from *n*-hexane - ethyl ether (1 : 1) to give pale yellow needles, mp 170 °C. The compound (7) was identified 7 by mixed melting point experiment and comparing the ¹H nmr and ir spectral data.

6-Prenylapigenin (9)

Compound (9) was recrystallized from *n*-hexane - ethyl ether (1 : 1) to give yellow needles, mp 223 °C. HR-ms: m/z 338.1127 (M⁺, C₂₀H₁₈O₅ requires 338.1149). ¹H Nmr(acetone- d_6 , 400 MHz): δ 1.65, 1.78 (each 3H, br s), 3.35 (2H, br d, J = 7 Hz), 5.28 (1H, m), 6.61, 6.62 (each 1H, s), 7.02 (2H, d, J = 9 Hz), 7.92 (2H, d, J = 9 Hz), 13.30 (1H, s). ¹³C Nmr (acetone- d_6 , 100 MHz): δ 161.9 (C-2), 104.1 (C-3), 183.2 (C-4), 105.3 (C-4a), 162.5 (C-5), 112.4 (C-6), 164.9 (C-7), 94.1 (C-8), 156.6 (C-8a), 123.5 (C-1'), 129.2 (C-2'and 6'), 116.9 (C-3' and 5'), 160.3 (C-4'), 20.0 (C-9), 123.3 (C-10), 131.7 (C-11), 25.9 (C-12), 17.9 (C-13).

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